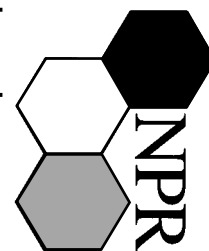


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Covering: 1995

Previous review: 1996, 13, 151

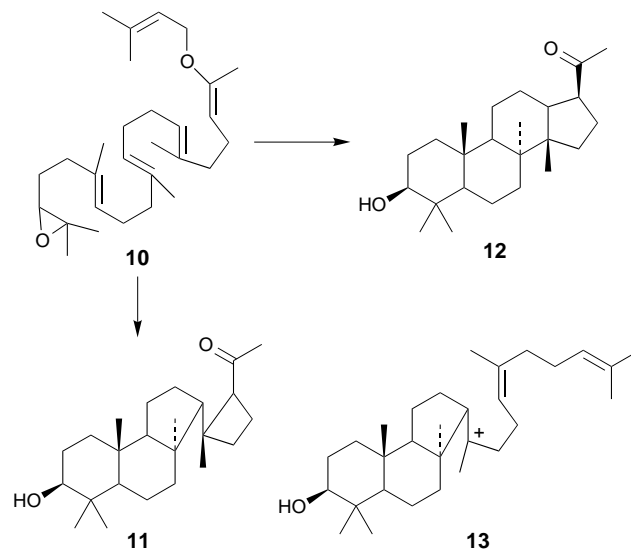
- 1 Introduction
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- 3 The fusidane–lanostane group
- 4 The dammarane–euphane group
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## 1 Introduction

This article follows the pattern of the previous report. The use of triterpenoids as biomarkers in terrestrial and marine sediments,<sup>1</sup> the pharmacology of oleanolic and ursolic acids<sup>2</sup> and the biological activity of triterpenoids which show sweetness inhibition<sup>3</sup> have been reviewed.

## 2 The squalene group

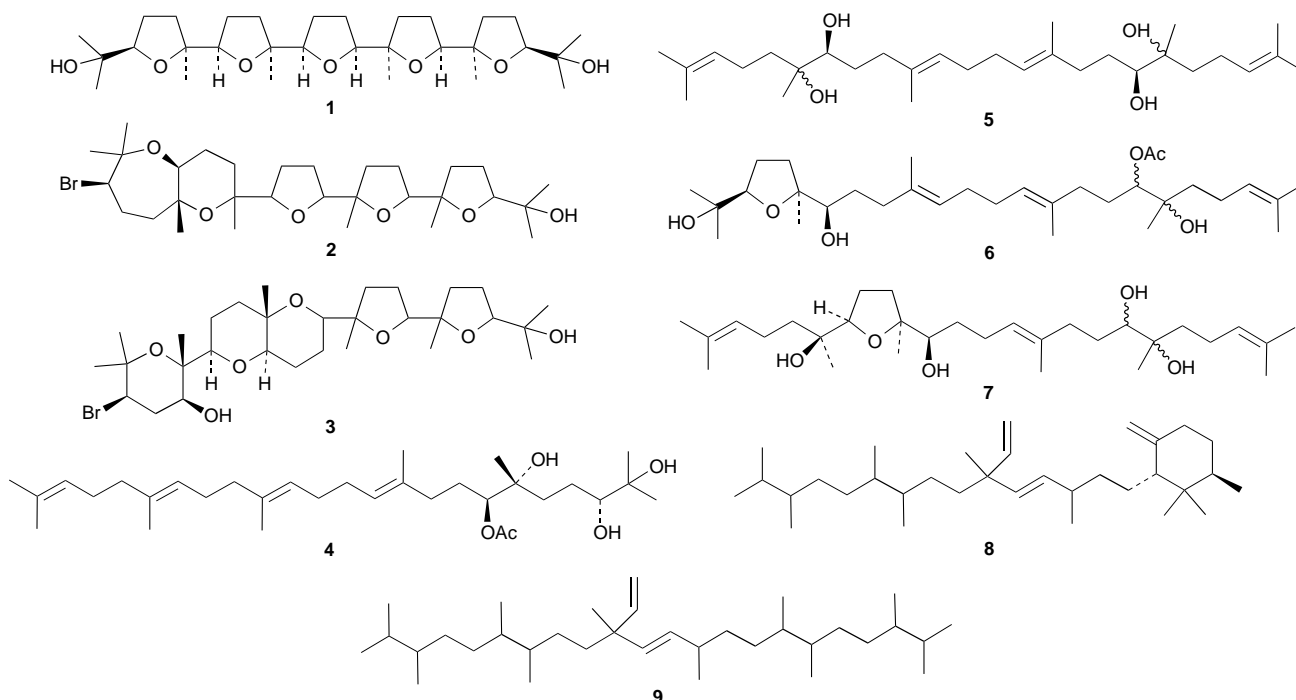
Glabrescol **1** is an interesting symmetrical squalene polytetrahydrofuran derivative from *Spathelia glabrescens*.<sup>4</sup> It is presumably derived from a squalene hexaepoxide. Enshuol **2** from *Laurencia omaezakiana*<sup>5</sup> and callicladol **3** from *L. calliclada*<sup>6</sup> must also arise from polyepoxysqualene precursors. Sapelin D **4** is a further oxygenated squalene from the bark of *Entandrophragma cylindricum*<sup>7</sup> (see *Nat. Prod. Rep.*, 1995, **12**, 609). A



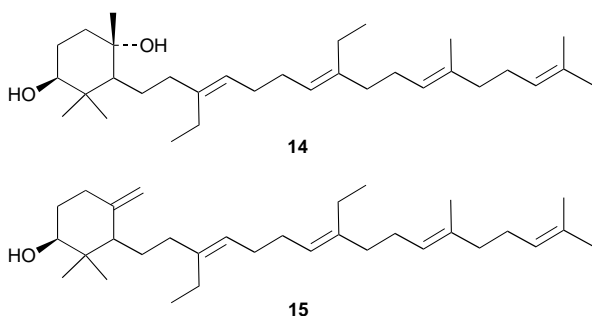
series of oxygenated squalenes has been isolated from *Quassia multiflora*.<sup>8</sup> Quassiol D **5** is a tetrol while quassiols B **6** and C **7** contain a tetrahydrofuran ring.

Sacridicene **8**, a monocyclic botryococcene, has been isolated from a lacustrine sediment deposited 10 280 years ago in Sacred Lake, Mount Kenya.<sup>9</sup> Octrahydrobotryococcene **9** has also been reported from sediments.<sup>10</sup>

The generally accepted concerted nature of the cyclisation of 2,3-oxidosqualene to lanosterol by lanosterol synthase from

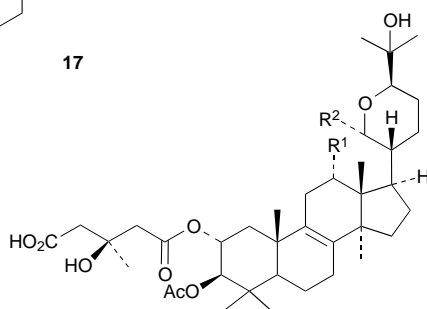
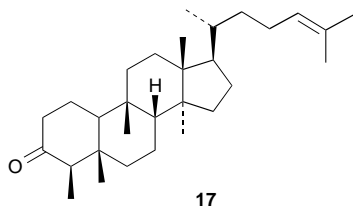
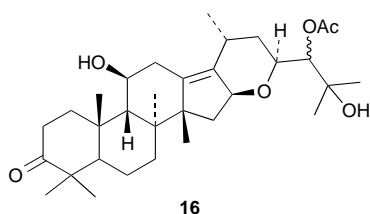


yeast has been challenged by Corey and his colleagues.<sup>11</sup> They argue that the formation of the minor product **11**, together with the expected product **12**, in the cyclisation of 20-oxa-2,3-oxidosqualene **10** suggests the intermediacy of discrete cations, like **13**, in the cyclisation of 2,3-oxidosqualene. (*S*)-2,3-Oxidosqualene has been prepared<sup>12</sup> from the corresponding 2-chloro-3-ketone by reduction with (*S*)-BINAL-H. Each of the (6*R*,7*R*)-, (6*S*,7*S*)-, (10*R*,11*R*)- and (10*S*,11*S*)-oxidosqualenes has been prepared and its absolute configurations determined.<sup>13</sup> Substitution of the methyl groups at C-10 and C-15 of 2,3-oxidosqualene with ethyl groups interferes with the normal cyclisation by the cyclase enzyme.<sup>14</sup> The reaction stops at the monocyclic stage with the formation of **14** and **15**. The former is a bis-homo achilleol A (see *Nat. Prod. Rep.*, 1994, **11**, 111).



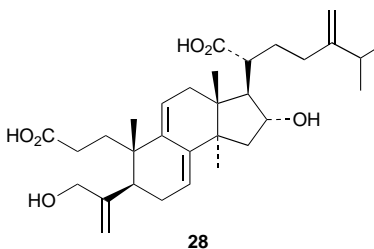
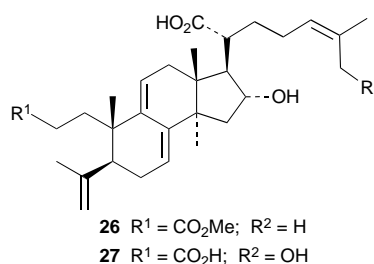
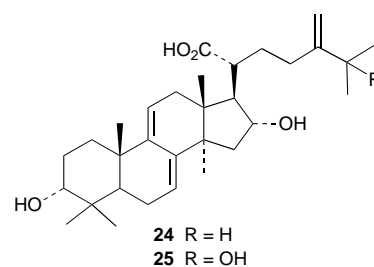
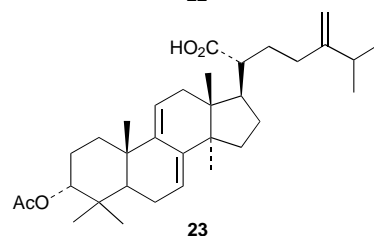
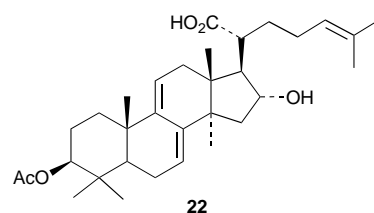
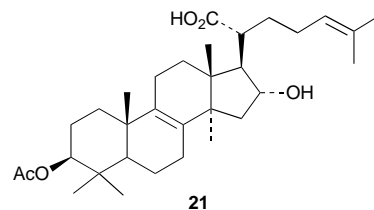
### 3 The fusidane-lanostane group

Alizexol A **16** is a novel protostane triterpenoid from the rhizomes of *Alisma orientalis*.<sup>15</sup> A rearranged lanostane, nigrum-24-en-3-one **17**, has been isolated from *Empetrum nigrum*<sup>16</sup> (the authors use a different numbering system, *i.e.*

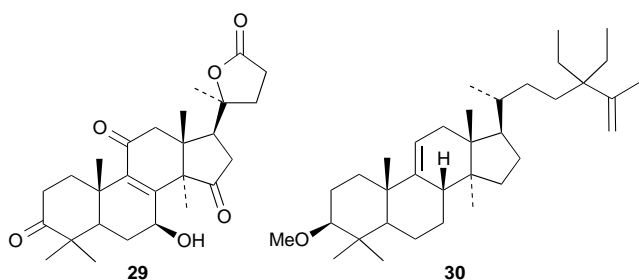


- 18** R<sup>1</sup> = OAc; R<sup>2</sup> = OH  
**19** R<sup>1</sup> = H; R<sup>2</sup> = OH  
**20** R<sup>1</sup> = H; R<sup>2</sup> = OAc

nigrum-21-en-3-one). Several crustinol derivatives, hebelomic acids **B 18**, **E 19** and **F 20**, have been isolated from the mushroom *Hebeloma senescens*.<sup>17</sup> *Poria cocos* contains the trametenolic acid and dehydrotrametenolic acid derivatives **21**, **22** and **23**.<sup>18</sup> Other lanostanes from the same source include 3-epidehydrotumulosic acid **24**, the corresponding 25-hydroxy derivative **25**, and the ring A cleaved compounds poricoic acids

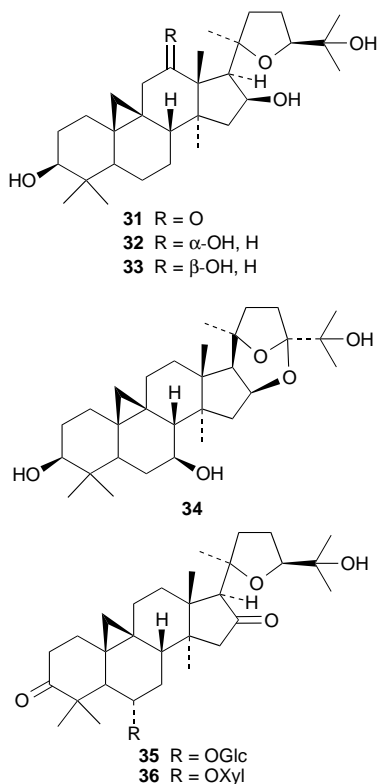


**B** (as the methyl ester) **26**, **E 27** and **F 28**.<sup>19</sup> The highly oxygenated gonolactone **A 29** is a metabolite of *Ganoderma lucidum*.<sup>20</sup> The 24,24-diethylhanostane *O*-methyllaureolol **30** has been isolated from *Skimmia laureola* subsp. *multinervia*.<sup>21</sup>

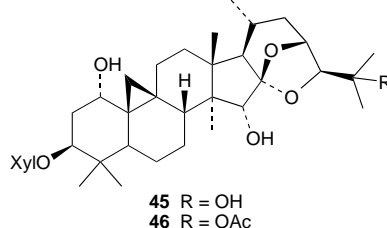
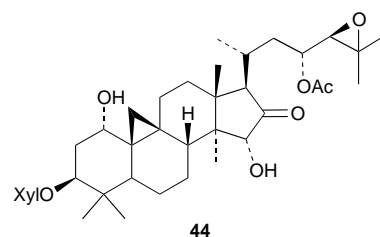
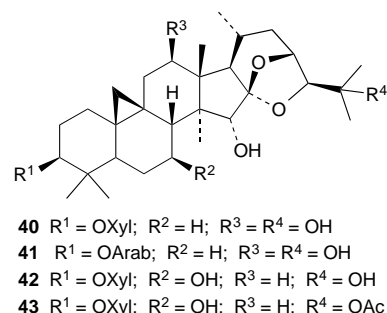
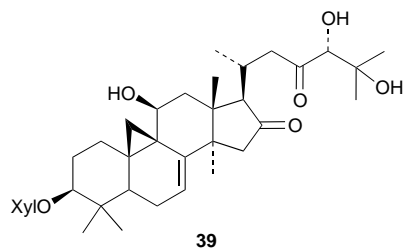
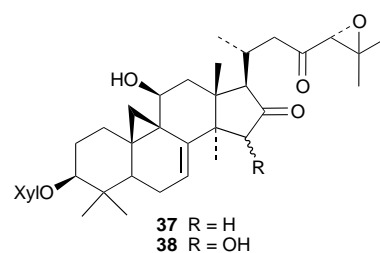


Experiments have shown that cycloartenol is not a precursor of lanosterol in *Euphorbia lathyris*<sup>22</sup> while parkeol [lanost-9(11)-en-3 $\beta$ -ol] is a precursor of the holostane saponins in the sea cucumbers *Holothura florida* and *Actinopygia agassiza*.<sup>23</sup>

Three closely related cycloartanes, cycloalpigenins A **31**, B **32** and C **33** and their corresponding 3-*O*- $\beta$ -D-xylopyranosides, cycloalpiosides A, B and C, have been isolated from *Astragalus alopecurus*.<sup>24, 25, 26</sup> Cycloalpigenin **34** and cycloalpioside, the corresponding 3 $\beta$ -D-xylopyranoside, are also present in *A. alopecurus*.<sup>27</sup> Two new glycosides **35** and **36** have been reported from *A. adsurgens*.<sup>28</sup> The saponins of several *Astragalus* species, *A. taschkendicus* (askendoside F)<sup>29</sup> *A. membranaceus* (astragalosides III and IV),<sup>30</sup> *A. trigonus* (trigonosides I–III),<sup>31</sup> *A. dissectus*, *A. ephemerotorum* and *A. kulabensis*,<sup>32</sup> have been reported.

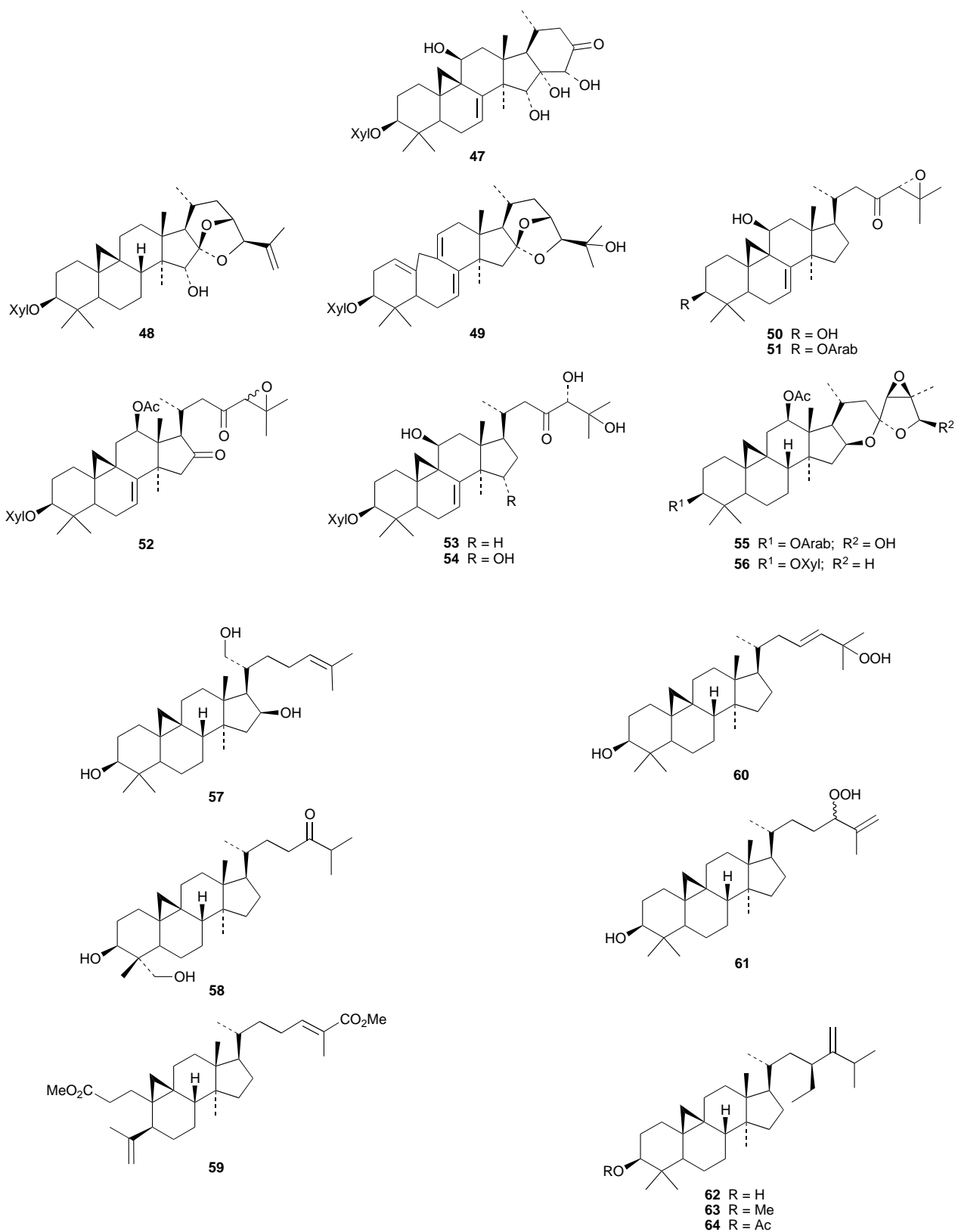


Cimicifugosides H<sub>1</sub> **37**, H<sub>2</sub> **38** and H<sub>5</sub> **39** are new saponins from *Cimicifuga simplex*.<sup>33</sup> The genins are all new and that of cimicifugoside H<sub>1</sub> **37** was confirmed by X-ray analysis of the corresponding diacetate. Four new cimigenol derivatives **40–43** have been found in *C. simplex*<sup>34</sup> which also contains the 1 $\alpha$ -hydroxyshengmanol derivative **44** and two further cimigenol derivatives **45** and **46**.<sup>35</sup> Full details of the isolation of cimicifugosides H<sub>3</sub> and H<sub>4</sub> (see *Nat. Prod. Rep.*, 1996, **13**, 153) have been reported<sup>36</sup> together with the new cimicifugoside H<sub>6</sub> **47**. Cimicifugosides H<sub>4</sub> and H<sub>6</sub> **47** have also been found in *C.*



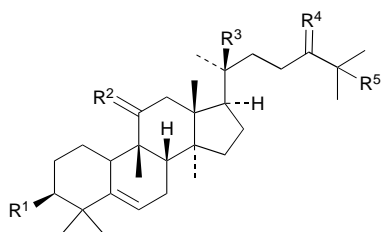
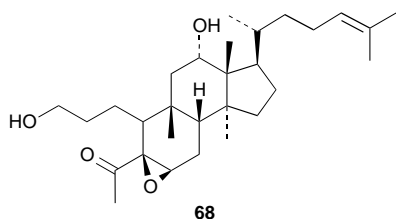
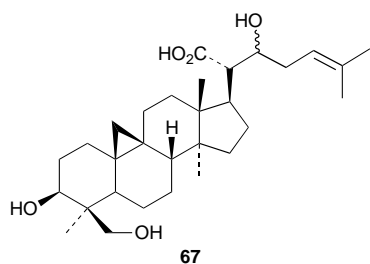
*foetida*<sup>37</sup> together with 25-anhydrocimigenol 3-*O*- $\beta$ -D-xyloside **48**, cimicinol 3-*O*- $\beta$ -D-xyloside **49**, cimicidanol **50**, cimicidanol 3-*O*- $\alpha$ -L-arabinoside **51**, cimicifol 3-*O*- $\beta$ -D-xyloside **52**, cimicidol 3-*O*- $\beta$ -D-xyloside **53** and the corresponding 15 $\alpha$ -hydroxy derivative **54**, and the actein derivative **55**. Cimigenol 3-*O*- $\beta$ -D-glucoside, cimicide F, also occurs in *C. foetida*.<sup>38</sup> The rhizomes of *Souliea vaginata* contain a range of cycloartane saponins, including a new compound, 26-deoxyactein **56**.<sup>39</sup>

Other cycloartanes include cycloart-24-ene-3 $\beta$ ,16 $\beta$ ,21-triol **57** from *Piptostygma fugax*,<sup>40</sup> 3 $\beta$ ,28-dihydroxycycloartan-24-one **58** from *Aglaia harmsiana*<sup>41</sup> and the dimethyl ester of nigranoic acid **59** from *Tillandsia usneoides*.<sup>42</sup> The hydroperoxides **60** and **61** have been reported from *Tillandsia recurvata*.<sup>43</sup> *Murraya exotica* contains<sup>44</sup> the side-chain

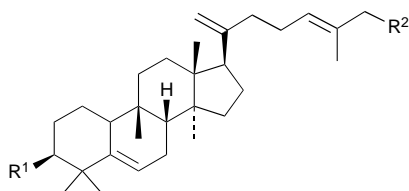


alkylated derivatives **62–66**. Thalictosides V and IX, from *Thalictrum* species, are based on the new genin **67**.<sup>45</sup> Mussaendoside F from *Mussaenda pubescens*<sup>46</sup> is a further saponin based on heinsigenin A (see *Nat. Prod. Rep.*, 1994, **11**, 96).

Lovenone **68** is an interesting degraded cucurbitane derivative from *Adalaria loveni*.<sup>47</sup> *Caput nigri* is a rich source of cucurbitane glycosides. Capenosides D–L **69–77** have been isolated and characterised.<sup>48, 49</sup> The pharmacological effects of cucurbitacins have been reviewed.<sup>50</sup>



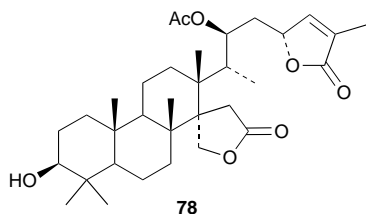
- 69 R<sup>1</sup> = GlcO; R<sup>2</sup> = O; R<sup>3</sup> = H; R<sup>4</sup> = OH,H; R<sup>5</sup> = OH  
 70 R<sup>1</sup> = GlcO; R<sup>2</sup> = α-OH,H; R<sup>3</sup> = H; R<sup>4</sup> = O; R<sup>5</sup> = GlcO  
 71 R<sup>1</sup> = Glc(1-2)GlcO; R<sup>2</sup> = α-OH,H; R<sup>3</sup> = H; R<sup>4</sup> = O; R<sup>5</sup> = GlcO  
 72 R<sup>1</sup> = Glc(1-6)GlcO; R<sup>2</sup> = α-OH,H; R<sup>3</sup> = H; R<sup>4</sup> = O; R<sup>5</sup> = GlcO  
 73 R<sup>1</sup> = Glc(1-6)GlcO; R<sup>2</sup> = O; R<sup>3</sup> = H; R<sup>4</sup> = O; R<sup>5</sup> = GlcO  
 74 R<sup>1</sup> = GlcO; R<sup>2</sup> = α-OH,H; R<sup>3</sup> = OH; R<sup>4</sup> = OH,H; R<sup>5</sup> = GlcO  
 75 R<sup>1</sup> = Glc(1-2)GlcO; R<sup>2</sup> = α-OH,H; R<sup>3</sup> = OH; R<sup>4</sup> = OH,H; R<sup>5</sup> = GlcO



- 76 R<sup>1</sup> = Glc(1-6)GlcO; R<sup>2</sup> = GlcO  
 77 R<sup>1</sup> = Glc(1-6)GlcO; R<sup>2</sup> = Glc(1-6)GlcO

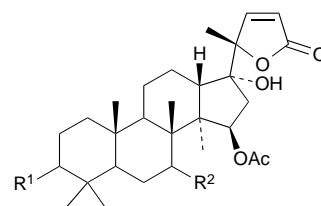
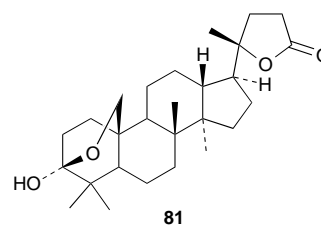
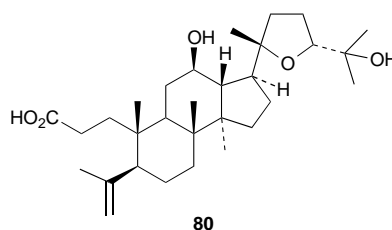
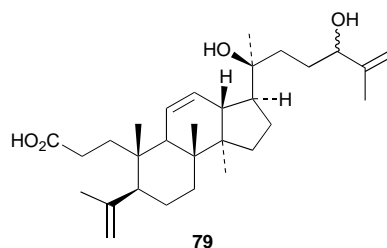
#### 4 The dammarane-euphane group

Hovenidulcigenin A **78** is the genin of hovenidulciosides A<sub>1</sub> and A<sub>2</sub> from *Hovenia dulcis*.<sup>51</sup> The structure of hovenidulcigenin A **78** was confirmed by X-ray analysis and shown to be



a 16,17-secodammarane with a methyl migration from C-20 to C-17. Cyclocariosides II and III from *Cyclocarya paliurus* are the 24-O-α-L-arabinopyranoside of the 3,4-secodammarane

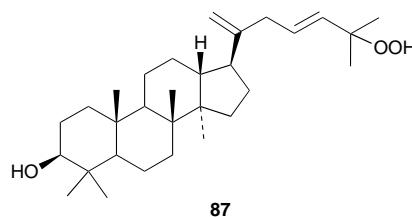
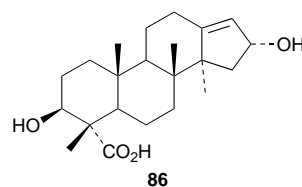
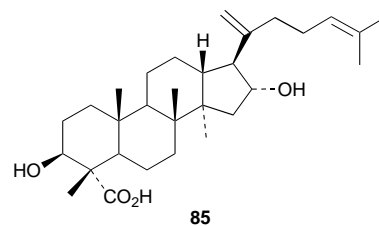
**79** and the 12-O-β-D-quinovopyranoside of the 3,4-secodammarane **80**, respectively.<sup>52</sup> Several 25,26,27-trinordammaranes have been found in *Cleome amblyocarpa*,<sup>53</sup> they have

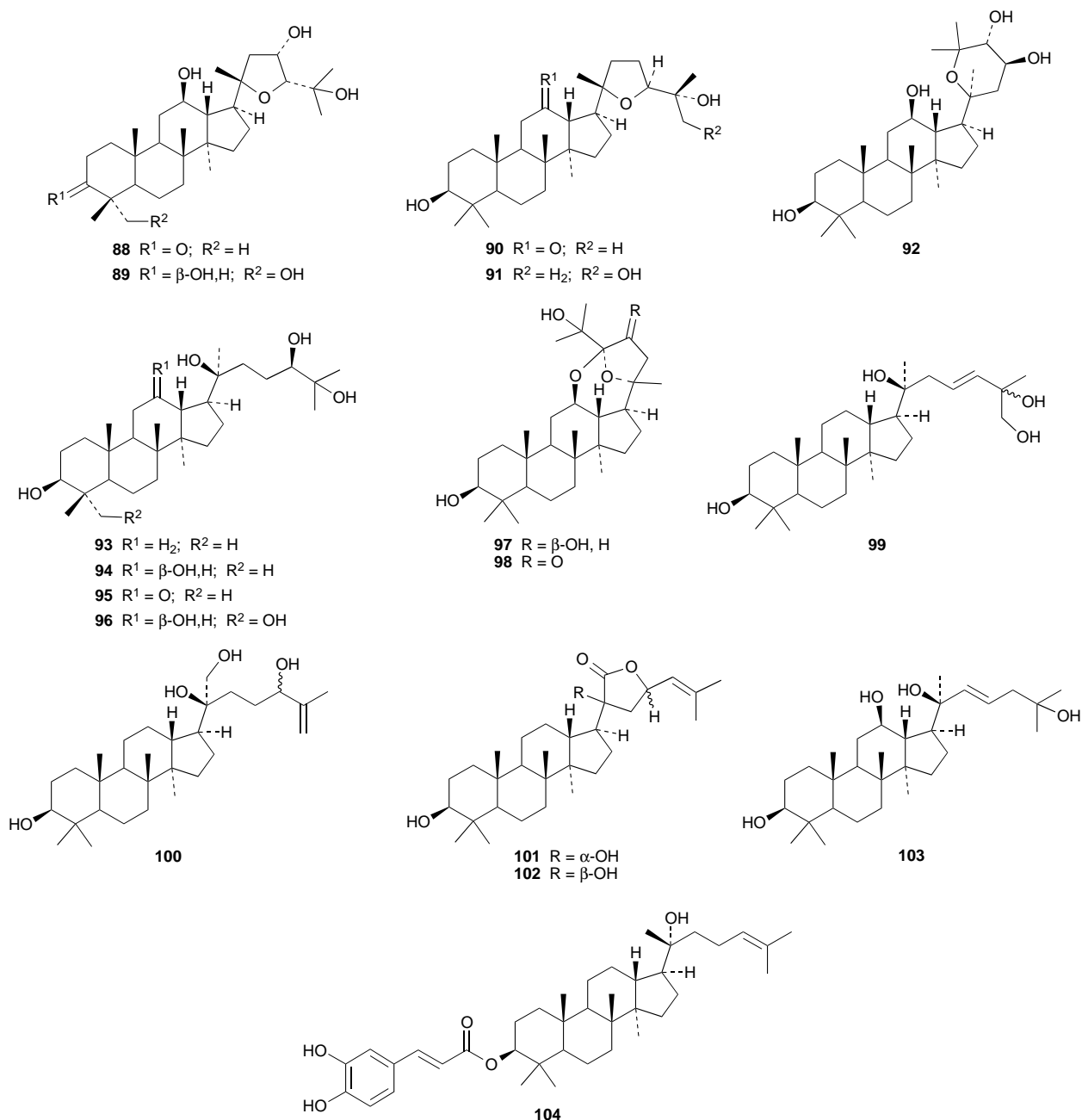


- 82 R<sup>1</sup> = β-OAc; R<sup>2</sup> = H

- 83 R<sup>1</sup> = R<sup>2</sup> = β-OAc

- 84 R<sup>1</sup> = α-OAc; R<sup>2</sup> = H



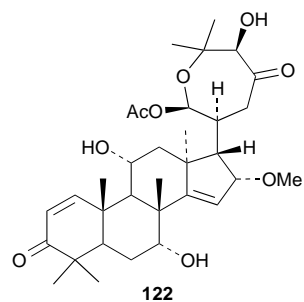
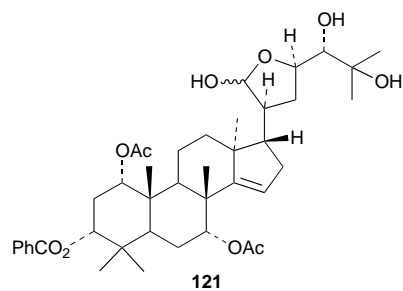
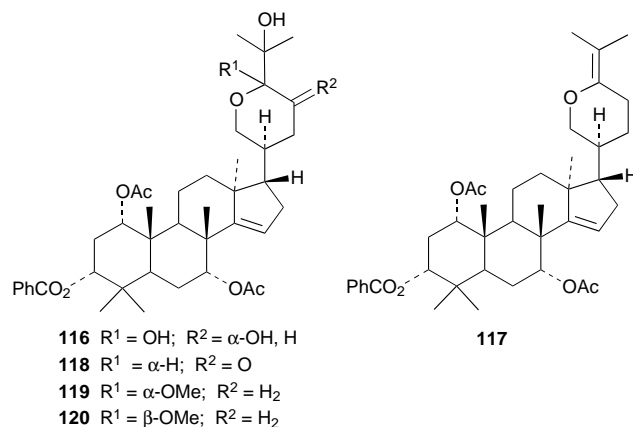
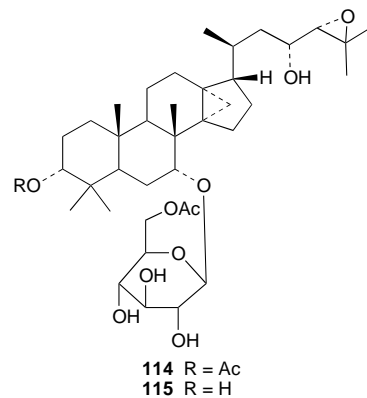
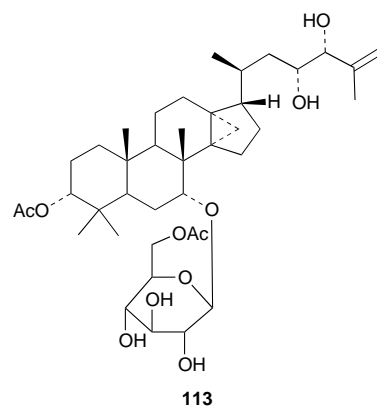
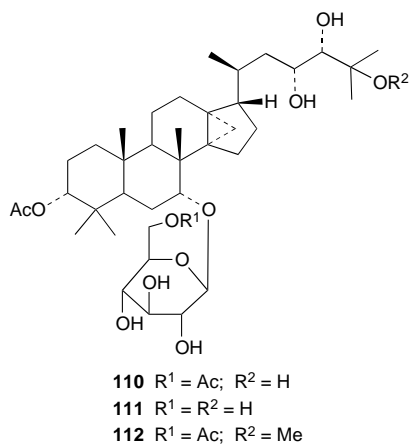
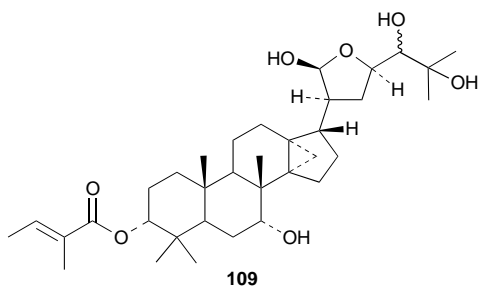
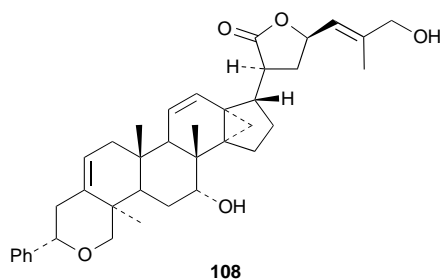
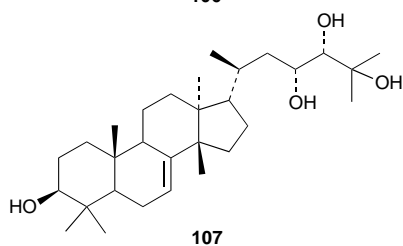
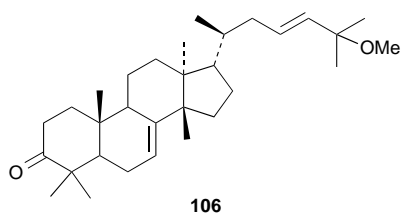
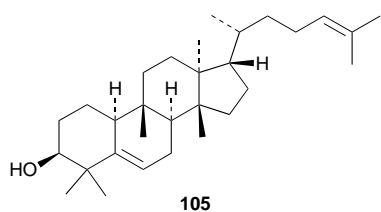


been named amblyone **81**, cleomblynols A **82** and B **83** and isocleomblynol A **84**. Rotundifolic acid from *Combretum rotundifolium* is the 16-*O*- $\alpha$ -L-arabinopyranoside of the new genin 3 $\beta$ ,16 $\alpha$ -dihydroxydammarane-20,24-dien-28-oic acid **85**.<sup>54</sup> The 16-*O*- $\alpha$ -L-arabinopyranoside of the nordammarane 3 $\beta$ ,16 $\alpha$ -dihydroxymansumbin-13(17)-en-28-oic acid **86** is also found in the same plant. 25-Hydroperoxydammarane-20,23-dien-3 $\beta$ -ol **87** has been isolated from *Boronia algida*.<sup>55</sup>

*Neosalsmitra integrifoliola* has produced 23 new saponins, the neoalsosides which include the new genins neolalsogenins B **88**, C **89**, G **90**, H **91**, M **92**, (20*S*,24*R*)-dammarane-3 $\beta$ ,20,24,25-tetrol **93**, (20*S*,24*R*)-dammarane-3 $\beta$ ,12 $\beta$ ,20,24,25-pentol **94** and its 12-ketone **95**, (20*S*,24*R*)-dammarane-3 $\beta$ ,12 $\beta$ ,20,24,25,28-hexol **96** and the acetals **97** and **98**.<sup>56, 57</sup> The 20,24-diols **93**–**96** all readily cyclised to the corresponding 20,24-epoxides on acid treatment. Four saponins from *Gynostemma pentaphyllum* have the new genins dammar-23-ene-3 $\beta$ ,20,25,26-tetrol **99**, dammar-25-ene-3 $\beta$ ,20,21,24-tetrol **100** and the epimeric lactones **101** and **102**.<sup>58</sup> Koryoginsenosides  $R_1$  and  $R_2$  from *Panax ginseng* include the new genin dammar-

22-ene-3 $\beta$ ,12 $\beta$ ,20,25-tetrol **103**.<sup>59</sup> Four dammarane saponins have been found in *Betula ermanii* together with the caffeate **104** of dammarenediol II.<sup>60</sup> Zizyotin is a saponin of jujubogenin from *Zizyphus oenoplea*<sup>61</sup> and another saponin of jujubogenin has been reported from *Centrosema bracteosum*.<sup>62</sup> Other new dammarane saponins with known genins include cyclocarioside I from *Cyclocarya paliurus*<sup>63</sup> and yixinoside A from *Gynostemma yixingense*.<sup>64</sup>

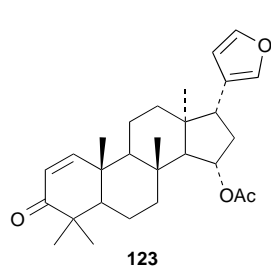
Boeticol **105** from *Euphorbia boetica* is 19(10 $\rightarrow$ 9)-abeo-eupha-5,24-dien-3 $\beta$ -ol.<sup>65</sup> The new tirucallane derivative leucophyllone **106** has been reported from *Aglaia leucophylla*,<sup>66</sup> and piscidinol B from *Walsura piscidia* has been shown to be identical with hispidol B **107**.<sup>67</sup> Dichapetalin A **108** is a glabretal analogue from *Dichapetalum madagascariense* with an unusual alkylation at C-3.<sup>68</sup> Another glabretal analogue **109** has been obtained from *Quassia multiflora*.<sup>69</sup> An X-ray analysis of cumingianoside A **110** has established the absolute configurations of cumingianosides A **110**, B, **111**, C **112**, D **113**, E **114** and F **115** from *Dysoxylum cumingianum*.<sup>70</sup>



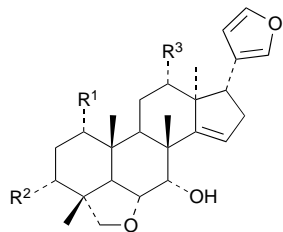
Several new apotirucallanes have been isolated from *Melia volkensii* including meliavolin **116**,<sup>71</sup> meliavolen **117**, melianone **118**,<sup>72</sup> meliavolkensins A **119** and B **120**,<sup>73</sup> and meliavolkenin **121**.<sup>74</sup> The structure of meliavolin **116** and the known melianin A<sup>72</sup> were confirmed by X-ray analysis. Piscicidol F **122** has been isolated from *Walsuria piscidia*.<sup>67</sup>

#### 4.1 Tetranortriterpenoids

Structure **123** has been proposed for neeflene, a tetranortriterpenoid from the dried flowers of *Azadirachta indica*.<sup>75</sup> The lack of an oxygen substituent at C-7 is unusual. Trichilinins B **124**



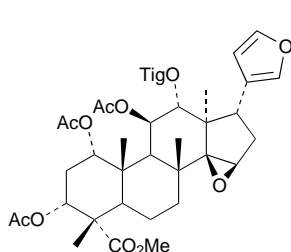
**123**



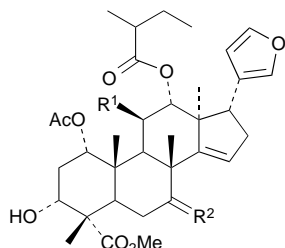
**124** R<sup>1</sup> = OTig; R<sup>2</sup> = R<sup>3</sup> = OAc

**125** R<sup>1</sup> = OAc; R<sup>2</sup> = OTig; R<sup>3</sup> = H

**126** R<sup>1</sup> = OCin; R<sup>2</sup> = OAc; R<sup>3</sup> = H



**127**



**128** R<sup>1</sup> = OAc; R<sup>2</sup> =  $\alpha$ -OAc, H

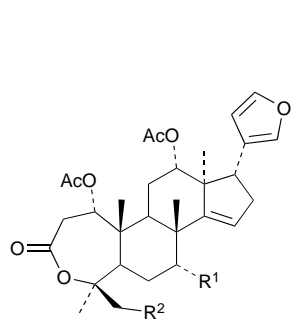
**129** R<sup>1</sup> = OH; R<sup>2</sup> =  $\alpha$ -OH, H

**130** R<sup>1</sup> = OAc; R<sup>2</sup> =  $\alpha$ -OH, H

**131** R<sup>1</sup> = OAc; R<sup>2</sup> = O

and C **125** are constituents of *Melia toosendan*<sup>76</sup> and meliavolkin **126** is found in *M. volkensii*.<sup>71</sup> Further examples of the highly oxygenated tetranortriterpenoids from *Turraea* species have been reported. *T. nilotica* yielded nilotin **127**<sup>77</sup> while *T. floribunda* gave compounds **128–131**.<sup>78</sup>

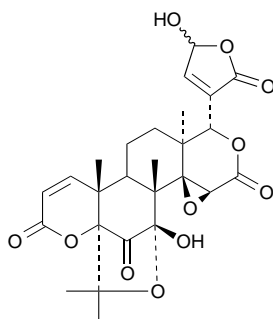
New ring-A cleaved tetranortriterpenoids include rubralins A **132**, B **133** and C **134** from *Trichilia rubra*<sup>79</sup> and



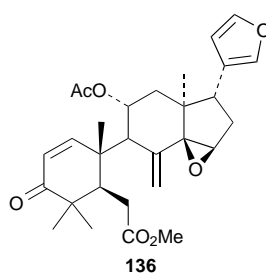
**132** R<sup>1</sup> = R<sup>2</sup> =  $\text{O}_2\text{C}-\text{CH}(\text{OH})-\text{CH}_2\text{CH}_3$

**133** R<sup>1</sup> =  $\text{O}_2\text{C}-\text{CH}(\text{OH})-\text{CH}_2\text{CH}_3$ ; R<sup>2</sup> =  $\text{O}_2\text{C}-\text{CH}(\text{OH})-\text{CH}_2\text{CH}_3$

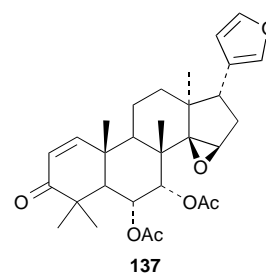
**134** R<sup>1</sup> =  $\text{O}_2\text{C}-\text{CH}=\text{CH}_2$ ; R<sup>2</sup> = H



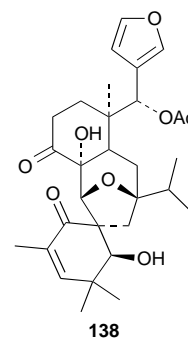
**135**



**136**



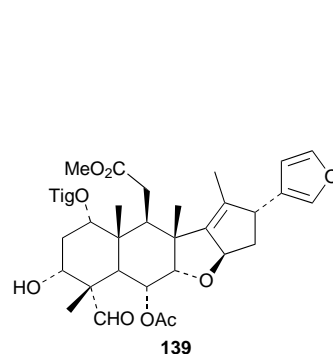
**137**



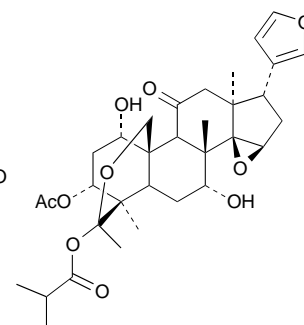
**138**

perforatinolone **135** from *Harrisonia perforata*.<sup>80</sup> 12-Deacetoxytoonacilin **136** occurs in *Toona ciliata* with the azadirone derivative **137**.<sup>81</sup> Entilin D **138** is a further example of the highly cleaved compounds from *Entandrophragma utile*.<sup>82</sup>

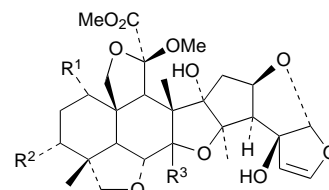
Further reports on the synthesis of azadirachtin<sup>83</sup> and the biologically active compounds from neem<sup>84</sup> have been



**139**



**140**



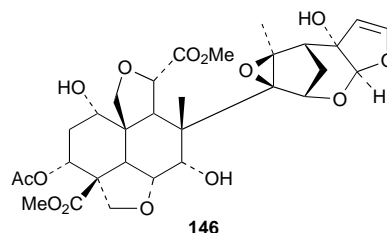
**141** R<sup>1</sup> = OTig; R<sup>2</sup> = OAc; R<sup>3</sup> =  $\alpha$ -H

**142** R<sup>1</sup> = OAc; R<sup>2</sup> = OTig; R<sup>3</sup> =  $\alpha$ -H

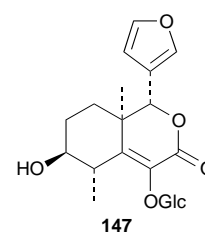
**143** R<sup>1</sup> = H; R<sup>2</sup> = OTig; R<sup>3</sup> =  $\beta$ -H

**144** R<sup>1</sup> = OAc; R<sup>2</sup> = OTig; R<sup>3</sup> =  $\beta$ -H

**145** R<sup>1</sup> = OTig; R<sup>2</sup> = OAc; R<sup>3</sup> =  $\beta$ -H



**146**



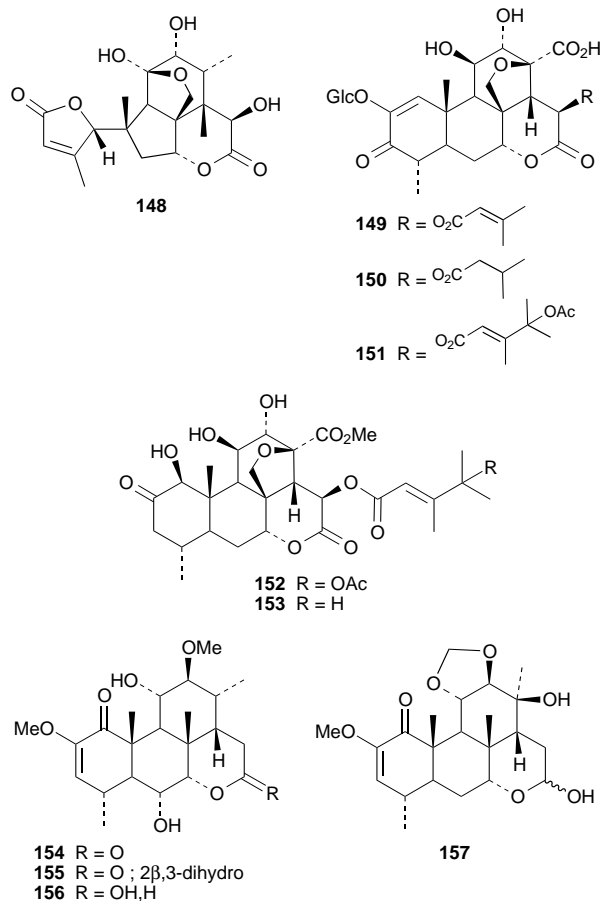
**147**



published. The flow of compounds from *Melia azedarach* continues. The latest additions are salannal **139**,<sup>85</sup> azedirachin C **140**,<sup>86</sup> and the meliacarpinin derivatives **141** and **142**,<sup>87</sup> **143–145**<sup>88</sup> and the 1-desacylazadirachtin derivative **146**.<sup>89</sup> Fagaropsin **147** is a degraded limonoid from *Fagaropsis glabra*.<sup>90</sup>

## 4.2 Quassinoids

Polyandrol **148** is a C-19 quassinoid from *Castela polyandra*.<sup>91</sup> Other new quassinoids include bruceosides D **149**, E **150** and

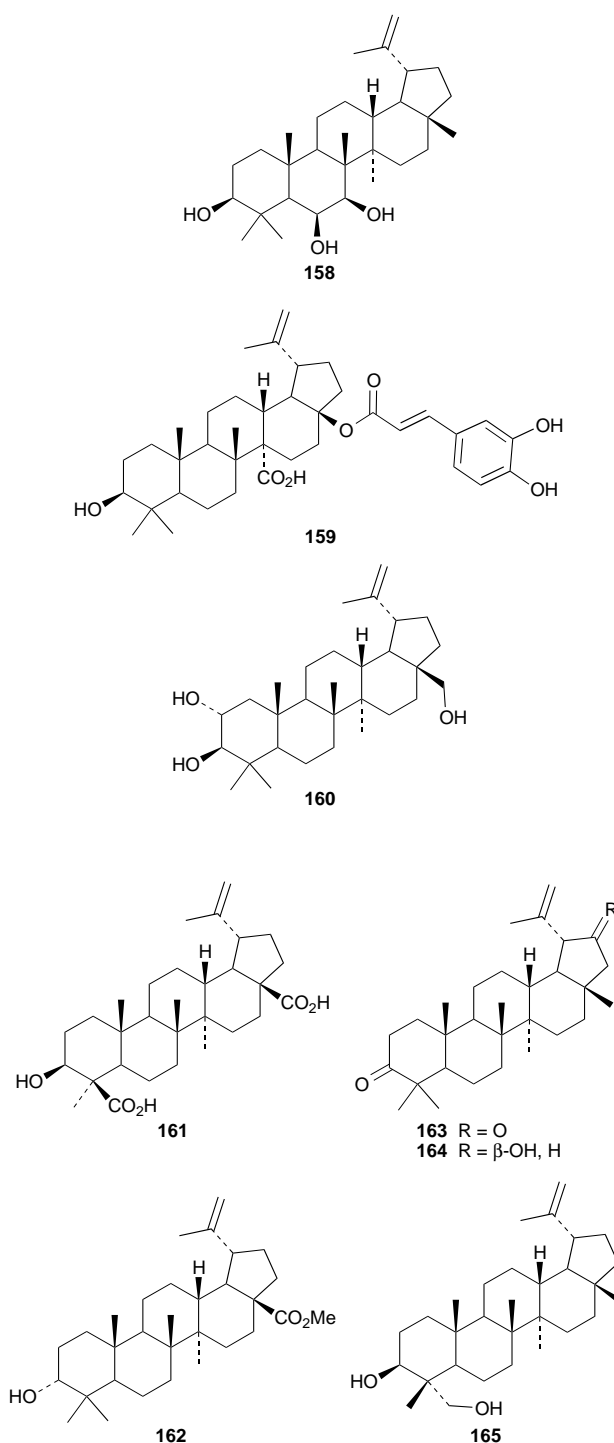


F **151** from *Brucea javanica*,<sup>92</sup> bruceanols G **152** and F **153** from *Brucea antidiysenterica*,<sup>93</sup> and javanicin Z **154**, dihydro-javanicin Z **155** and hemiacetaljavanicin Z **156** from *Picrasma javanica*.<sup>94</sup> The structure of **156** was confirmed by X-ray analysis. Picrasinol D **157** is a constituent of *Picrasma ailanthoides*.<sup>95</sup>

## 5 The lupane group

Two new lupane derivatives, the 3 $\beta$ ,6 $\beta$ ,7 $\beta$ -triol **158** and the rare 27-carboxylic acid **159**, have been isolated from *Rhoiptelea chiliantha*.<sup>96</sup> The 2 $\alpha$ ,3 $\beta$ ,28-triol **160** and the corresponding 3,28-dipalmitate have been found in *Rheum rhabarbarium*.<sup>97</sup> Other new lupanes include 3 $\beta$ -hydroxylup-20(29)-ene-24,28-dioic acid **161** from *Mimusops elengi*,<sup>98</sup> methyl 3-*epi*-betulinate **162** from *Syzygium samarangense*,<sup>99</sup> salacianone **163** and salacianol **164** from *Salacia beddomei*,<sup>100</sup> lup-20(29)-ene-3 $\beta$ ,23-diol **165** from *Bursera simaruba*<sup>101</sup> and lupeol caffeate from *Betula ermanii*.<sup>60</sup>

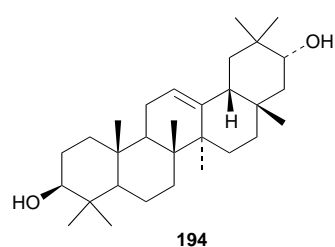
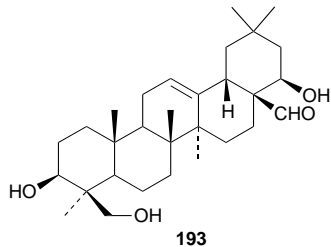
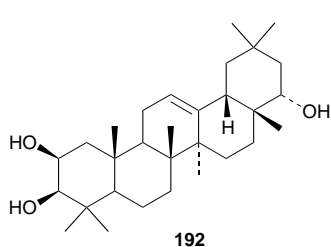
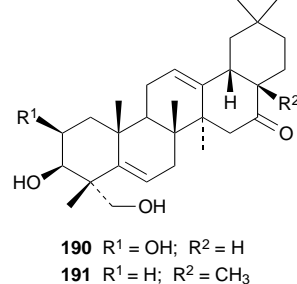
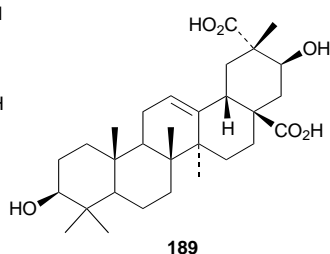
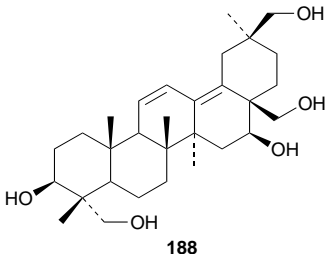
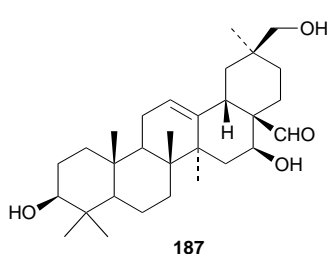
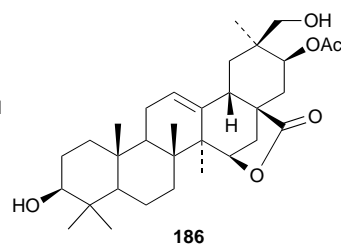
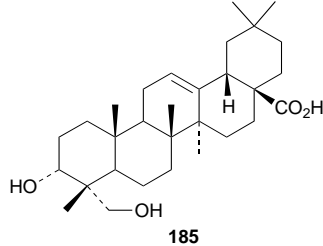
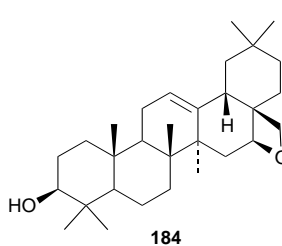
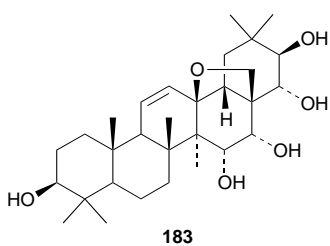
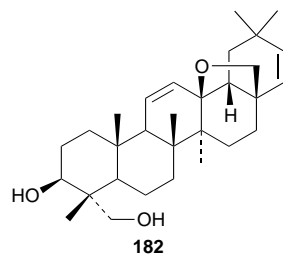
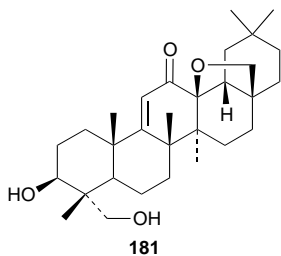
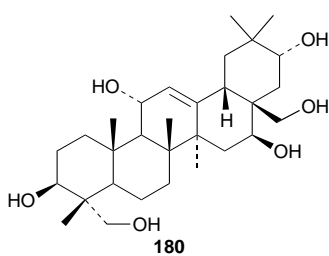
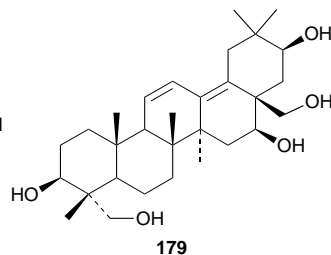
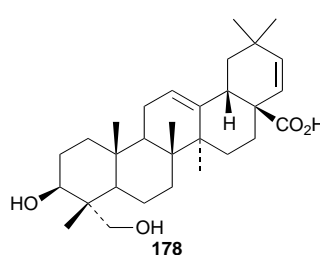
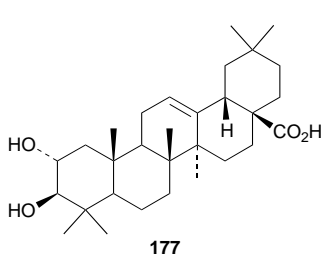
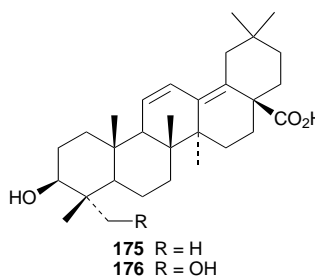
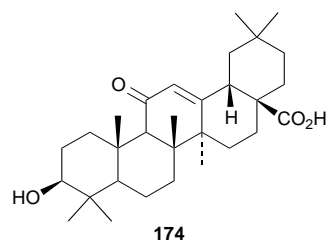
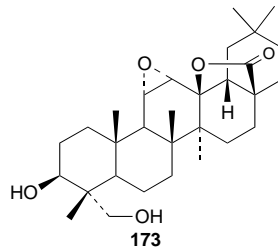
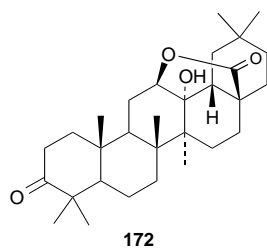
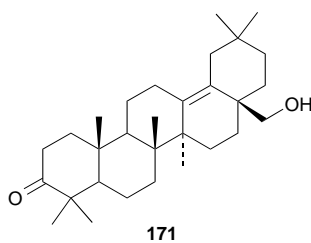
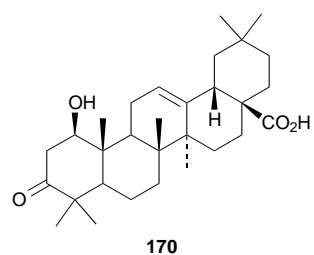
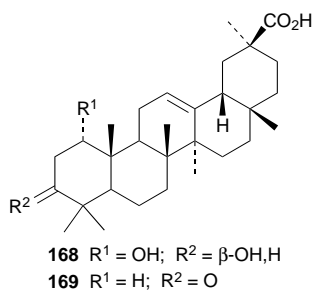
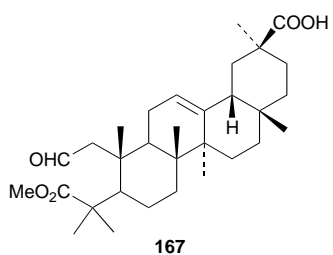
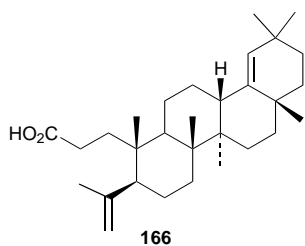
Three lupane glycosyl esters, oplopanaxosides B, C and D, have been isolated from the leaves of *Oplopanax japonicus*.<sup>102</sup> and have been shown to be saponins of 3 $\beta$ -hydroxylup-20(29)-ene-23,28-dioic acid, 3 $\alpha$ ,23-dihydroxylup-20(29)-en-28-oic acid



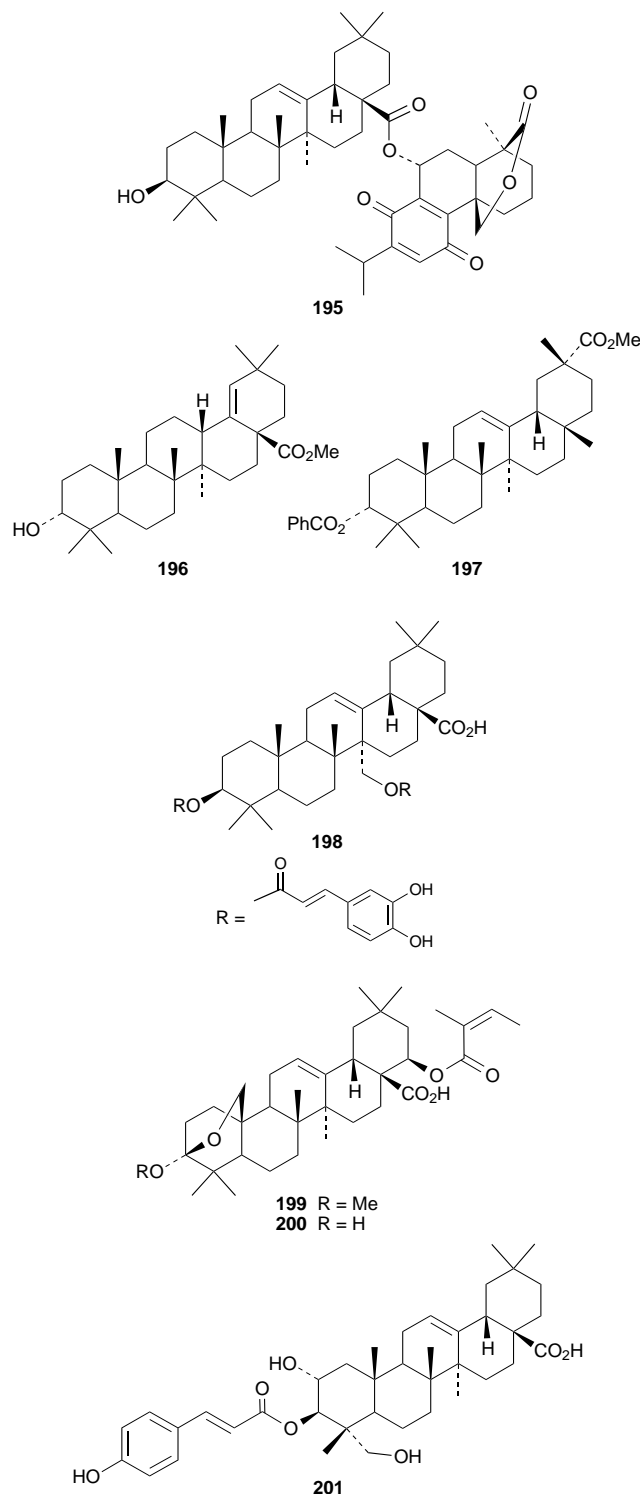
and 3 $\beta$ ,23-dihydroxylup-20(29)-en-28-oic acid, respectively. These saponins have previously been published as the cirenso-sides F, H and G, respectively.<sup>103</sup> A lupane saponin has been reported from *Coccinia indica*.<sup>104</sup>

## 6 The oleanane group

3,4-Secooleana-4(23),18-dien-3-oic acid **166** has been found in *Vahlia capensis*<sup>105</sup> while the 2,3-secooleanane dillenic acid D **167** has been isolated from *Dillenia papuana* together with dillenic acid E **168** and 3-oxoolean-12-en-30-oic acid **169**.<sup>106</sup> 1 $\beta$ -Hydroxy-3-oxoolean-12-en-28-oic acid **170** has been detected in *Limnophila rugosa* and 28-hydroxyolean-13(18)-en-3-one **171**, named sabianone, has been found in *Sabinia*

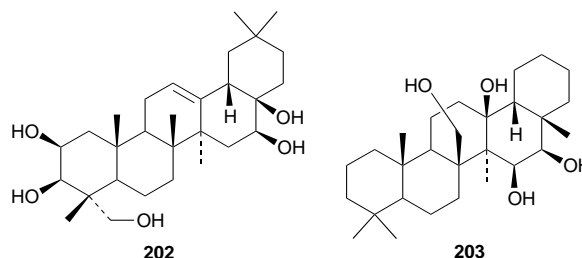


*swinhoei*, together with sabialactone **172**.<sup>107</sup> *Paeonia japonica* is the source of 3 $\beta$ ,23-dihydroxy-11 $\alpha$ ,12 $\alpha$ -epoxyoleanan-28,13 $\beta$ -olide **173**, 3 $\beta$ -hydroxy-11-oxoolean-12-en-28-oic acid **174** and 3 $\beta$ -hydroxyoleana-11,13(18)-dien-28-oic acid **175** whereas the corresponding 23-hydroxy derivative **176** and 2 $\alpha$ ,3 $\beta$ -dihydroxyolean-12-en-28-oic **177** are found in *Paeonia suffruticosa*.<sup>108</sup> The clinopodisides B–G are saponins from *Clinopodium chinensis* with the new sapogenins 3 $\beta$ ,23-dihydroxyoleana-12,21-dien-28-oic acid **178**, oleana-11,13(18)-diene-3 $\beta$ ,16 $\beta$ ,21 $\beta$ ,23,28-pentol **179**, olean-12-ene 3 $\beta$ ,11 $\alpha$ ,16 $\beta$ ,21 $\alpha$ ,23,28-hexol **180**, the 13 $\beta$ ,28-epoxyoleananes **181** and **182**.<sup>109</sup> 13 $\beta$ ,28-Epoxyolean-11-ene-3 $\beta$ ,15 $\alpha$ ,16 $\alpha$ ,21 $\beta$ ,22 $\alpha$ -pentol **183** is the genin of the new saponin ranuncoside VII from

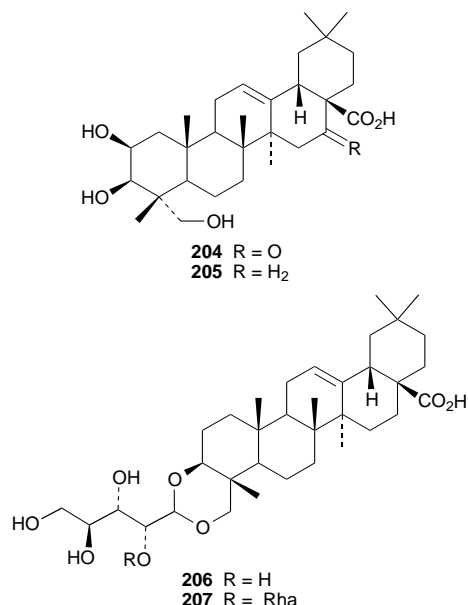


*Hydrocotyle ranunculoides*.<sup>110</sup> Liguveitol **184**, the unusual 16 $\beta$ ,28-epoxyolean-12-en-3 $\beta$ -ol, is the genin of liguveitoside A from *Ligularia veitchiana*.<sup>111</sup> Other new oleananes include wilforol C **185**, the 3-epimer of hederagenin from *Tripterygium wilfordii*,<sup>112</sup> bridgesigenin C **186** from *Trichocereus pachanoi*,<sup>113</sup> alamosenogenin **187** from *Rathbunia alamosensis*,<sup>114</sup> oleana-11,13(18)-diene-3 $\beta$ ,16 $\beta$ ,23,28,30-pentol, saikaogenin Q **188**, the genin of saikosaponin Q from *Bupleurum smithii* va. *parvifolium*,<sup>115</sup> philoxeroic acid **189** from *Alternanthera philoxeroides*,<sup>116</sup> mimusopgenone **190** and mimugenone **191** from *Mimusops elengi*,<sup>117</sup> olean-12-ene-2 $\beta$ ,3 $\beta$ ,22 $\alpha$ -triol **192** from *Picris hieracioides*,<sup>118</sup> 3 $\beta$ ,22 $\beta$ ,24-trihydroxyolean-12-en-28-al **193**, the genin of lablalsaponin I from *Dolichos lablab*<sup>119</sup> and glyyunnansapogenin **194** from *Glycyrrhiza yunnanensis*.<sup>120</sup>

Reglin **195** from *Salvia regia* is an ester of oleanolic acid and the abietane deacetylsessein which both occur in the plant.<sup>121</sup> Other new naturally occurring oleanane esters include methyl 3 $\alpha$ -hydroxyolean-18-en-28-oate **196** from a *Frullania* sp.,<sup>122</sup> methyl 3-benzoylkatanoate **197** from *Limnophila heterophylla*,<sup>123</sup> the bis-caffeoyl ester **198** of 3 $\beta$ ,27-dihydroxyolean-12-en-28-oic acid from *Rhoiptelea chiliantha*,<sup>96</sup> camarilic acid **199**<sup>124</sup> and camaric acid **200**<sup>125</sup> from *Lantana camara* and 3-coumaroylarjunolic acid **201** from *Myrianthus liberecus*.<sup>126</sup>



Vicogenin **202** is a 28-noroleanane from *Vicoa indica*<sup>127</sup> and the unusual 29,30-dinoroleanane-13 $\beta$ ,15 $\beta$ ,16 $\beta$ ,26-tetrol **203** has been found in the soft coral *Nephthea albida*.<sup>128</sup> A full assignment of the <sup>1</sup>H and <sup>13</sup>C NMR spectra of the 27-noroleanane pyrocincholic acid has been reported.<sup>129</sup> Bernardioside B<sub>4</sub> is a new saponin from *Bellis bernardii* along with the new sapogenin bellisonic acid **204**.<sup>130</sup> The polygalasaponins XI–XIV from *Polygala japonica* include a new genin named polygalagenin **205**.<sup>131</sup> Anemocleomides A **206**



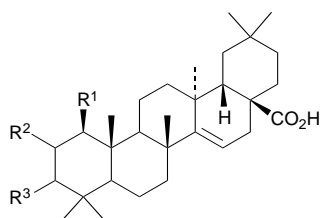
**Table 1** New oleanane saponins

Compound(s)	Source	Ref.
Achyranthosides C and D	<i>Achyranthes fauriei</i>	133
Albiziasaponins A, B and C	<i>Albizia lebbbeck</i>	134
Aradecoside	<i>Aralia decaisneana</i>	135
Armatoside	<i>Aralia armata</i>	136
Asterbatanosides D and E	<i>Aster batangensis</i>	137
Asterbatanosides F, G, H and I	<i>Aster batangensis</i>	138
Asterbat anosides J and K	<i>Aster batangensis</i>	139,140
Asteryunnanosides A, B, C and D	<i>Aster yunnanensis</i>	141,142
Asteryunnanoside E	<i>Aster yunnanensis</i>	143
Asteryunnanosides F and G	<i>Aster yunnanensis</i>	144
Asteryunnanoside H	<i>Aster yunnanensis</i>	145
Bellidiastroside C <sub>2</sub>	<i>Aster bellidiastrum</i>	146
Bersimosides I and II	<i>Trifolium alexandrinum</i>	147
Betavulgarosides I–V	<i>Beta vulgaris</i>	148
Cephalariasaponin B	<i>Cephalaria transsylvanica</i>	149
Clematichinensosides A and B	<i>Clematis chinensis</i>	150
Clematosides A–G	<i>Clematis stans</i>	151
Clemantoside E	<i>Clematis montana</i>	152
Elatosides G–K	<i>Aralia elata</i>	153
Esculentosides L <sub>1</sub> and R	<i>Phytolacca acinosa</i>	154
2'-O-β-D-Glucopyranosylmomordins Ic and IIc	<i>Kochia scoparia</i>	155
Glycoside L-6d	<i>Hedera helix</i>	156
Hacquetia saponins 1–4	<i>Hacquetia epipactis</i>	157
Herniaria saponin A	<i>Herniaria fontanesii</i>	158
Inflasaponins II and VI	<i>Glycyrrhiza inflata</i>	159
Japondipsaponin E <sub>1</sub>	<i>Dipsacus japonicus</i>	160
Kalopanaxsaponins G and H	<i>Kalopanax sepemlobus</i>	161
Kudzusaponins SA <sub>1</sub> –SA <sub>3</sub> and C <sub>1</sub>	<i>Pueraria lobata</i>	162
Linderniosides A and B	<i>Lindernia pyxidaria</i>	163
Lucyoside O	<i>Luffa cylindrica</i>	164
Lucyoside P	<i>Luffa cylindrica</i>	165
Mazusaponins I and III	<i>Mazus miquelii</i>	166
Melilotosides A–C	<i>Melilotus albus</i>	167
Melilotoside D	<i>Melilotus albus</i>	168
Mimusopin and Mimusopsin	<i>Mimusops elengi</i>	169
Mussaendoside S	<i>Mussaenda pubescens</i>	170
Patensin	<i>Pulsatilla patens</i> var. <i>multifida</i>	171
Polyandrasides A and B	<i>Phytolacca polyandra</i>	172
Polygalasaponins I–X	<i>Polygala japonica</i>	173
Reinosides A–F	<i>Polygala reinii</i>	174
Rubicunoside A	<i>Silene rubicunda</i>	175
Saikosapons M and N	<i>Bupleurum smithii</i>	176
Saponins E3, E6–E8	<i>Ilex dumosa</i>	177
(E)- and (Z)-Senegasaponins A and B, (Z)-Senegins II and III	<i>Polygala senega</i> var. <i>latifolia</i>	178,179
Sideroxylosides B and C	<i>Sideroxylon foetidissimum</i>	180
Songarosaponins E and F	<i>Verbascum songaricum</i>	181
Symphytoxide B	<i>Symphytum officinale</i>	182
Tauroside St-I <sub>2</sub>	<i>Hedera taurica</i>	183
Terminolitin	<i>Terminalia arjuna</i>	184
Thalicoside D	<i>Thalictrum minus</i>	185
Transsylvanosides E and F	<i>Cephalaria transsylvanica</i>	186
Trifosides A–C	<i>Akebia trifoliata</i>	187
Wistariasaponins A <sub>2</sub> , A <sub>3</sub> , B <sub>3</sub> , YC <sub>1</sub> and YC <sub>2</sub>	<i>Wisteria brachybotrys</i>	188
Yiyeliangwanosides VII and VIII	<i>Nothopanax davidii</i>	189
Yiyeliangwanosides IX–XI	<i>Nothopanax davidii</i>	190

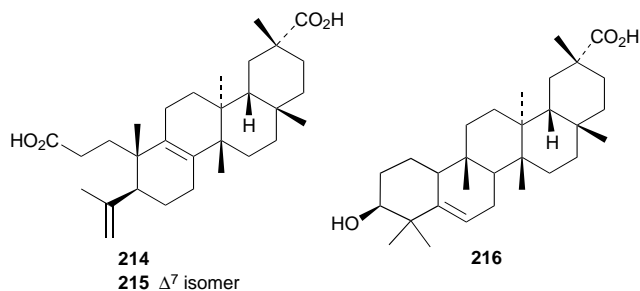
and B **207** are saponins from *Anemoclema glaucifolium* with an unusually linked arabinose.<sup>132</sup> Other named oleanane saponins with known genins are listed in Table 1.

The following plant sources have been reported to contain unnamed saponins of known oleanane genins: *Alalia armata*,<sup>191</sup> *Catunaregam nilotica*,<sup>192</sup> *Dumasia truncata*,<sup>193</sup> *Enterospermum pruinsum*,<sup>194</sup> *Gypsophila capillaris*,<sup>195</sup> *Gypsophila oldhamiana*,<sup>196</sup> *Mimosa pigra*,<sup>197</sup> *Myrsine australis*,<sup>198</sup> *Phytolacca rivinoides* and *Phytolacca bogotensis*,<sup>199</sup> *Pometia eximia*,<sup>200</sup> *Silene jennisensis*,<sup>201</sup> *Solidago virgaurea* subsp. *virgaurea*,<sup>202</sup> *Sophora japonica*<sup>203</sup> and *Vigna unguiculata* subsp. *unguiculata*.<sup>204</sup>

A host of taraxerane derivatives has been isolated from *Maprounea africana* and *M. membranacea*.<sup>205, 206</sup> These include the *p*-coumaroyl ester **208** of aleuritolic acid, the 2 $\alpha$ -hydroxy-**209** and 2 $\beta$ -hydroxy-3-epialeuritolic acid **210**, the 2,3-bis-(*p*-hydroxybenzoyl) ester of 1 $\beta$ ,2 $\alpha$ -dihydroxyaleuritolic acid **211** and the 3-(*p*-hydroxybenzoyl) esters of 2 $\alpha$ -hydroxyaleuritolic acid **212** and 1 $\beta$ -hydroxyaleuritolic acid **213**. The last named ester was previously thought to be an ester of 7 $\beta$ -hydroxy-maprounic acid. The X-ray crystal structure analysis of the known bryononic acid has been reported.<sup>207</sup> Bryononic acid was found in *Sandoricum koetjape* together with secobryononic acid **214** and the corresponding  $\Delta^7$ -isomer, secoisobryononic

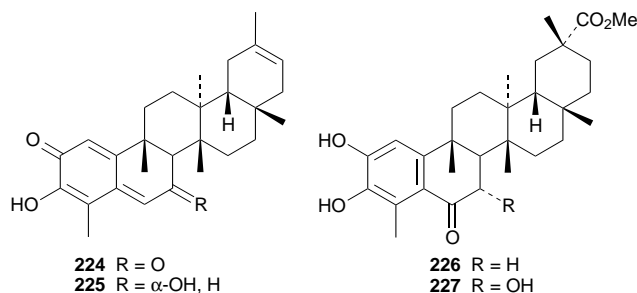
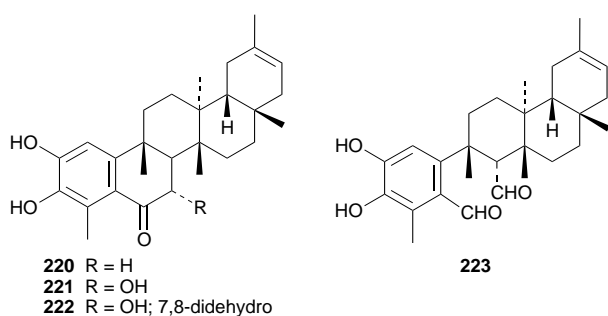
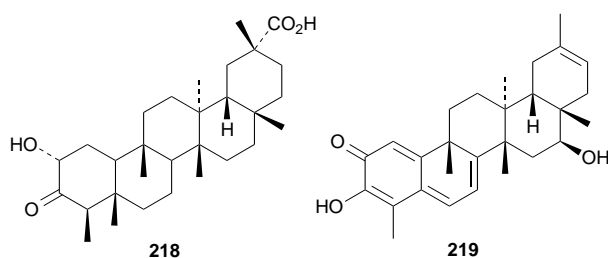
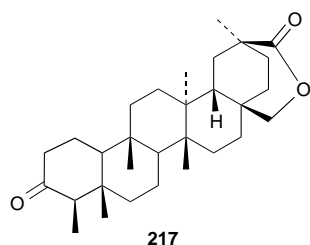


- 208**  $R^1 = R^2 = H$ ;  $R^3 = \beta$ -coumaroyloxy  
**209**  $R^1 = H$ ;  $R^3 = \beta$ -OH;  $R^2 = \alpha$ -*p*-hydroxybenzoyloxy  
**210**  $R^1 = H$ ;  $R^2 = \beta$ -*p*-hydroxybenzoyloxy;  $R^3 = \alpha$ -OH  
**211**  $R^1 = OH$ ;  $R^2 = \alpha$ -*p*-hydroxybenzoyloxy;  $R^3 = \beta$ -*p*-hydroxybenzoyloxy  
**212**  $R^1 = H$ ;  $R^2 = \alpha$ -OH;  $R^3 = \beta$ -*p*-hydroxybenzoyloxy  
**213**  $R^1 = OH$ ;  $R^2 = H$ ;  $R^3 = \beta$ -*p*-hydroxybenzoyloxy

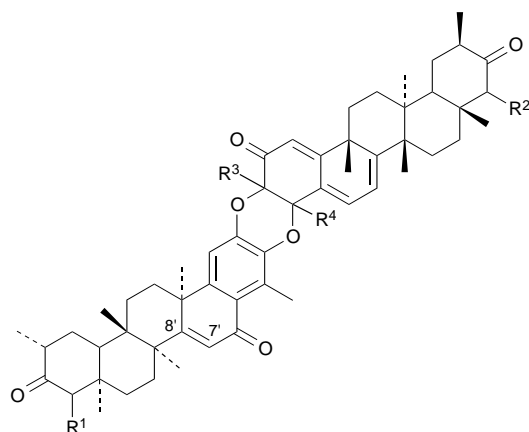


acid **215**. Wilforol D **216** is a glutinane derivative from *Trypterigium wilfordii*.<sup>112</sup>

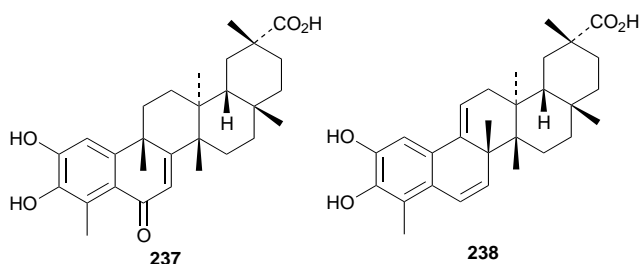
Several friedelane derivatives have been isolated from *Maytenus* species including maytenfolone **217** from *M. diversifolia*,<sup>208</sup> 2 $\alpha$ -hydroxy-3-oxofriedelan-29-oic acid **218**, 16 $\beta$ -hydroxyiguesterin **219**,<sup>209</sup> canarol **220**, 7 $\alpha$ -hydroxycanarol **221**, 7 $\alpha$ -hydroxy-6-oxo-7,8-didehydroiguesterol **222**, canarodial **223**, iguesteroquinone **224** and 7 $\alpha$ -hydroxy-7,8-



dihydroiguesterin **225** from *M. canariensis* and blepharodol **226** and 7 $\alpha$ -hydroxyblepharodol **227** from *M. blepharodes*.<sup>210</sup> The dimeric friedelane derivatives xuxuarine A $\alpha$  **228**, A $\beta$  **229**, Ba **230**, B $\beta$  **231**, Ca **232**, C $\beta$  **233**, Da **234** and D $\beta$  **235** and 7',8'-dihydroxuxuarine A $\beta$  **236** have been isolated from *M. chuchuhuasca*.<sup>211</sup> Wilforol A **237**, a norfriedelane derivative,

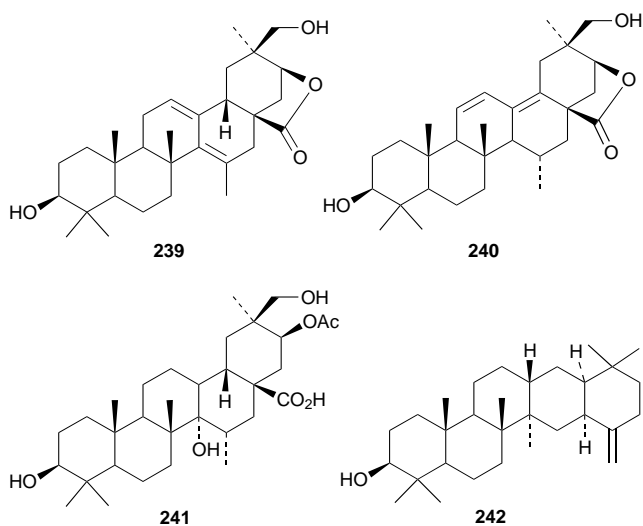


- 228**  $R^1 = R^2 = H$ ;  $R^3 = \beta$ -OH;  $R^4 = \beta$ -CH<sub>3</sub>  
**229**  $R^1 = R^2 = H$ ;  $R^3 = \alpha$ -OH;  $R^4 = \alpha$ -CH<sub>3</sub>  
**230**  $R^1 = \alpha$ -OH;  $R^2 = \beta$ -OH;  $R^3 = \beta$ -OH;  $R^4 = \beta$ -CH<sub>3</sub>  
**231**  $R^1 = \alpha$ -OH;  $R^2 = \beta$ -OH;  $R^3 = \alpha$ -OH;  $R^4 = \alpha$ -CH<sub>3</sub>  
**232**  $R^1 = H$ ;  $R^2 = \beta$ -OH;  $R^3 = \beta$ -OH;  $R^4 = \beta$ -CH<sub>3</sub>  
**233**  $R^1 = H$ ;  $R^2 = \beta$ -OH;  $R^3 = \alpha$ -OH;  $R^4 = \alpha$ -CH<sub>3</sub>  
**234**  $R^1 = \alpha$ -OH;  $R^2 = H$ ;  $R^3 = \beta$ -OH;  $R^4 = \beta$ -CH<sub>3</sub>  
**235**  $R^1 = \alpha$ -OH;  $R^2 = H$ ;  $R^3 = \alpha$ -OH;  $R^4 = \alpha$ -CH<sub>3</sub>  
**236**  $R^1 = R^2 = H$ ;  $R^3 = \alpha$ -OH;  $R^4 = \alpha$ -CH<sub>3</sub>; 7',8'-dihydro



and wilforol B **238**, the acid from isopristerin III, have been isolated from *Tripterigium wilfordii*.<sup>212</sup> The X-ray analysis of 7 $\beta$ ,8 $\beta$ -epoxyfriedelin has been published.<sup>213</sup> 7 $\beta$ ,8 $\beta$ -Epoxyfriedelin was derived from putranjivadiene (friedelane-3,7-dione).

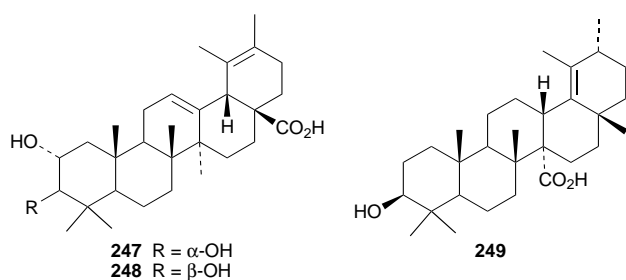
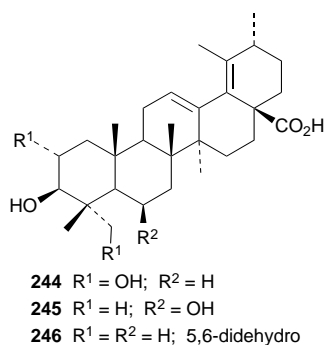
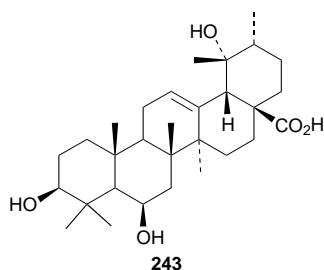
Pachanol A **239**, B **240** and C **241** are pachanane derivatives from *Trichocereus pachanoi*.<sup>113</sup> The structure of pachanol A **239** was confirmed by X-ray analysis. Ixerenol **242** is a rearranged oleanane from *Ixeris chinensis*.<sup>214</sup> Full <sup>1</sup>H and <sup>13</sup>C NMR



assignments of olean-18-ene, olean-13(18)-ene, olean-12-ene, taraxer-14-ene, multiflor-7-ene, multiflor-9(11)-ene, friedel-3-ene and friedelin have been reported.<sup>215</sup>

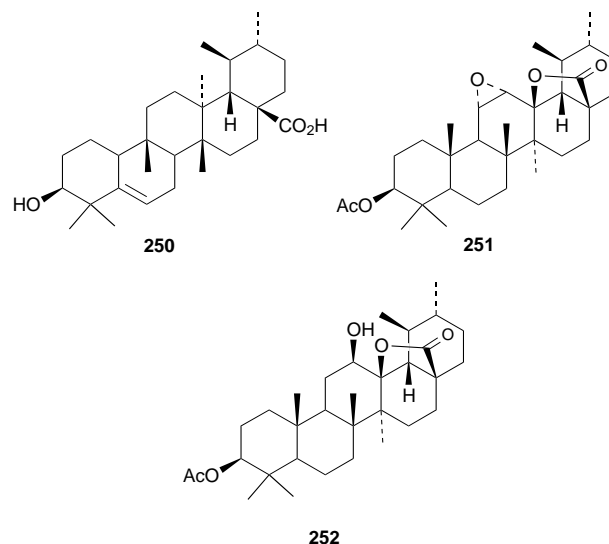
## 7 The ursane group

The structure of uncaric acid from *Uncaria thwaitesii* and *U. elliptica* has been revised to 3β,6β,19-trihydroxyurs-12-en-28-oic acid **243**.<sup>216</sup> It was formerly thought to be 3β,6β,18β-trihydroxyurs-12-en-30-oic acid. The structure of the 3β,6β-diacetate and the 3,6-diketone must also be revised. A sapogenin from *Rubus pinfaensis* has been shown to be 2α,3β,23-trihydroxyursa-12,18-dien-28-oic acid **244** by X-ray

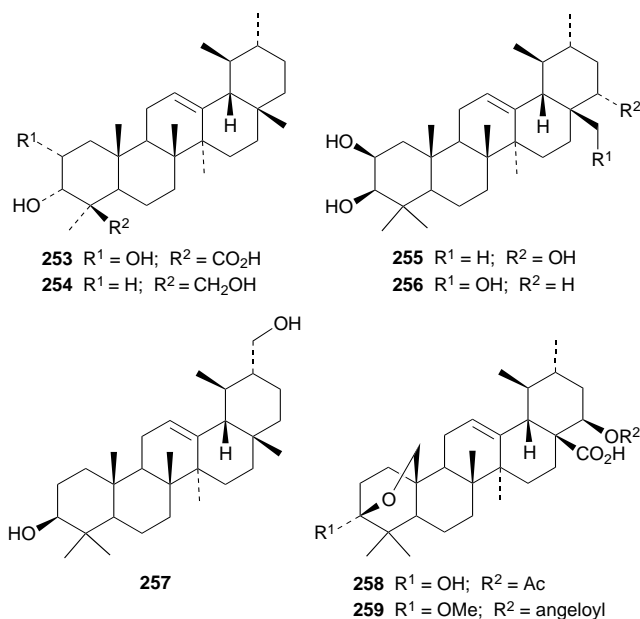


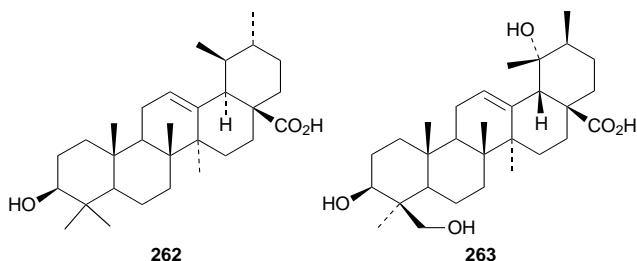
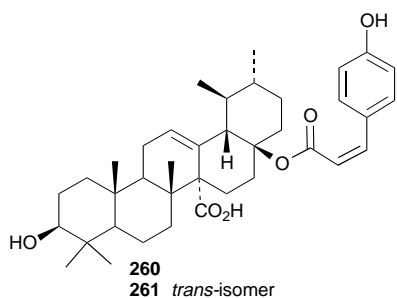
analysis of its triacetate methyl ester.<sup>217</sup> Two similar dienes, uncargenins A **245** and B **246** have been isolated from *U. rhynchophylla*.<sup>218</sup> They were accompanied by 3β,6β,23-trihydroxyolean-12-en-28-oic acid (uncargenin C) which has been isolated previously.<sup>219</sup> Two C-3 epimeric urs-12,19-dienes **247** and **248** have been found in *Rosa taiwanensis*.<sup>220</sup> 3β-Hydroxy-18-ursen-27-oic acid **249** occurs in *Adenocalymma alliaceum*.<sup>221</sup>

Rhoiptelic acid **250** is a rearranged ursane from *Rhoiptelea chiliantha*.<sup>222</sup> Two lactone derivatives **251** and **252** have



been isolated from *Bursera delpechiana*.<sup>223</sup> The deacetyl derivative of **251** is a known compound.<sup>224</sup> Other new ursanes include 2α,3α-dihydroxyurs-12-en-24-oic acid **253** and urs-12-ene-3α,24-diol **254** from *Boswellia serrata*,<sup>225</sup> urs-12-ene-2β,3β,22α-triol **255** and urs-12-ene-2β,3β,28-triol **256** from *Picris hieracioides*,<sup>118</sup> grahamidiol **257** from *Pristimera grahamii*,<sup>226</sup> camarinic acid **258**<sup>125</sup> and camaracinic acid **259**<sup>124</sup> from *Lantana camara*, *cis*-karenin **260** and *trans*-karenin **261** from *Nerium oleander*,<sup>227</sup> and 18α-ursolic acid, named as morinoursolic acid B **262**, from *Morina longifolia* together



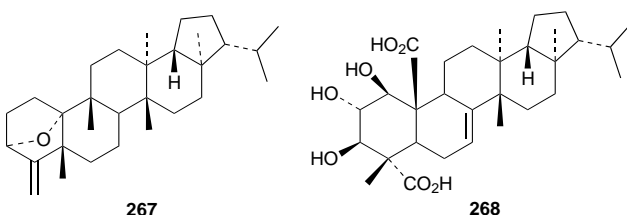
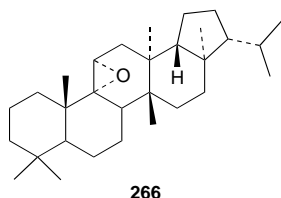
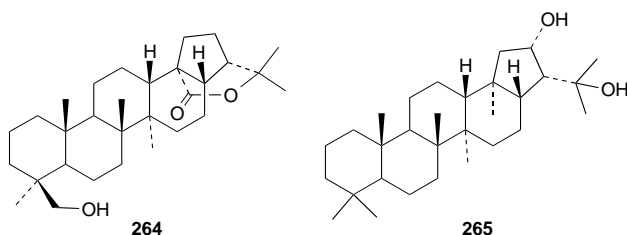


with the known 3-*epi*-ursolic acid, named as morinoursolic acid A.<sup>228</sup>

The genin of the main saponin ILA-1 of the leaves of *Ilex argentina* is claimed to be (20*S*)-3 $\beta$ ,19 $\alpha$ ,24-trihydroxyurs-12-en-28-oic acid **263**.<sup>229</sup> Ursane saponins have been reported from *Adina rubella* (rubellosides A and B),<sup>230</sup> *Blumea lacera*,<sup>231</sup> *Ilex paraguariensis*,<sup>232</sup> *Majorana hortensis*,<sup>233</sup> *Mazus miquelii* (mazusaponins II and IV),<sup>166</sup> *Mussaenda pubescens* (Mussaendoside R),<sup>170</sup> *Nauclea idierrichii*,<sup>234</sup> *Prunella vulgaris* (pruvulosides A and B)<sup>235</sup> and *Zygophyllum coccineum*, *Z. album* and *Z. dumosum* (zygophylloside F).<sup>236</sup>

## 8 The hopane group

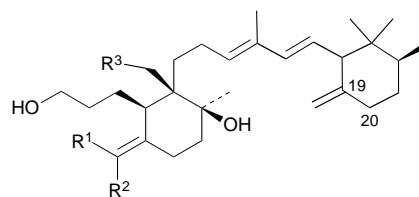
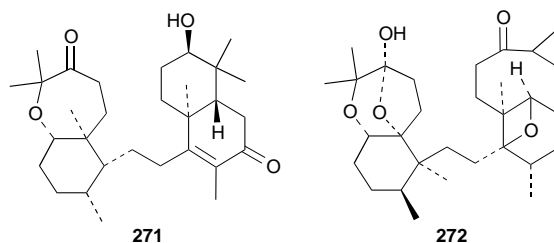
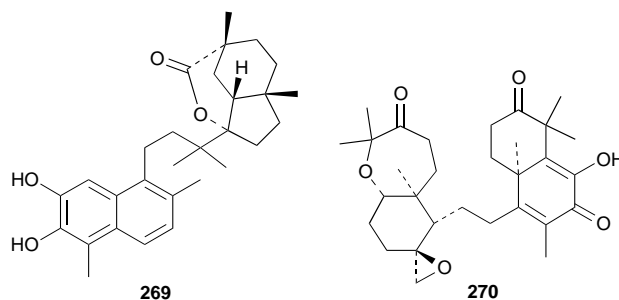
Diplaziosides I and II are hopane saponins from *Diplazium subsinuatum*.<sup>237</sup> The former has a new genin, 24-hydroxyhopan-28,22-olide **264**. Hopane-20 $\alpha$ ,22-diol **265** occurs in *Cyathia lepifera*<sup>238</sup> together with 9 $\alpha$ ,11 $\alpha$ -epoxyfernene **266**, the corresponding 9 $\beta$ ,11 $\beta$ -epoxide and



3 $\alpha$ ,10 $\alpha$ -epoxyficilic-4(23)-ene **267**. A highly oxygenated fern-7-ene derivative, polytolypin **268**, has been obtained from the coprophilous fungus *Polytolypa hystricis*.<sup>239</sup>

## 9 Miscellaneous compounds

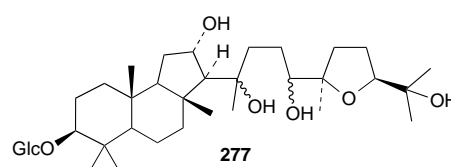
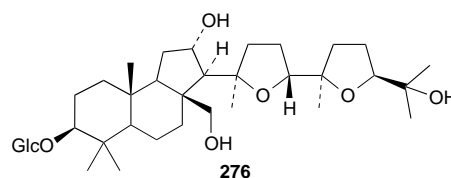
Tryptolactone **269** is an unusual C<sub>27</sub> compound from *Trypterigium wilfordii*.<sup>240</sup> Sodwanones G **270**, H **271** and I **272**, new members of the fascinating sipholane group of

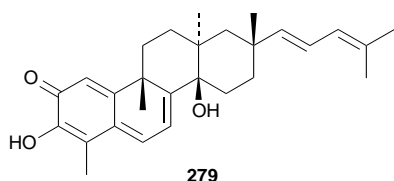
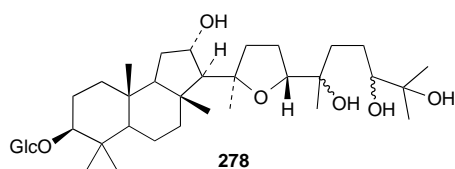


- 273** R<sup>1</sup> = CHO; R<sup>2</sup> = CH<sub>3</sub>; R<sup>3</sup> = H  
**274** R<sup>1</sup> = CH<sub>3</sub>; R<sup>2</sup> = CHO; R<sup>3</sup> = OH  
**275** R<sup>1</sup> = CH<sub>3</sub>; R<sup>2</sup> = CHO; R<sup>3</sup> = OH;  $\Delta^{19}$

triterpenoids, have been reported from *Axinella weltneri*.<sup>241</sup> The structure of sodwanone G **270** was confirmed by X-ray analysis, as were the structures of the previously reported sodwanones E and F. Three new compounds of the iridal group, irisgermanicals A **273**, B **274** and C **275**, have been isolated from *Iris germanica*.<sup>242</sup>

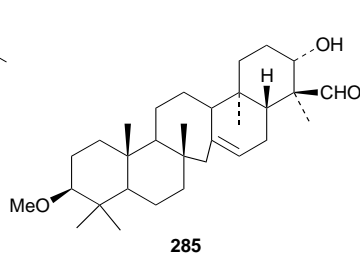
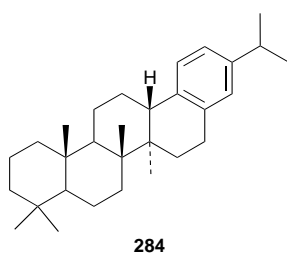
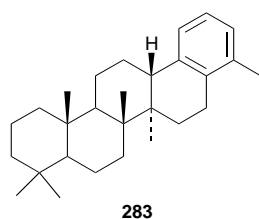
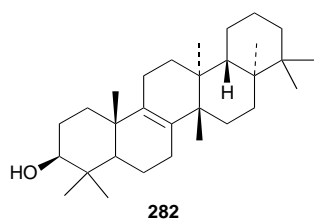
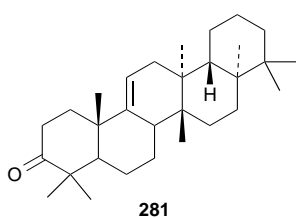
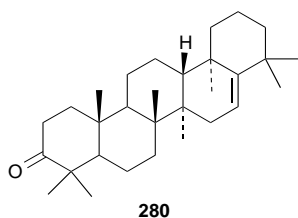
*Adesmia aconcaguensis* is the source of three new malabaricanes **276**, **277** and **278**.<sup>243</sup> A new shionane,





russulaflavidin **279**, has been found in the fungus *Russula flavida*.<sup>244</sup>

*Picris hieracioides* subsp. *japonicum* contains several 16-gammacerene and pichierene derivatives.<sup>245</sup> The ketone **280** is the only new gammacerene, the others having been previously described in 1989.<sup>246</sup> Pichierenone **281** and



isopichierenol **282** are also new although isopichierenyl acetate has been isolated previously. Some of the <sup>13</sup>C NMR spectroscopic assignments of members of both series have been modified. The ring D aromatic gammacerane derivatives **283** and **284** have been reported from Messel shale.<sup>247</sup> The serratene aldehyde **285** has been found in *Picea jezoensis*.<sup>248</sup>

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